

Association Of Genetic Variants Rs1888747 And Rs10868025 Of FRDM3 Gene With Diabetic Neuropathy In Pakistani Patients

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Abstract

Background: Diabetic kidney disease (DKD) may cause renal failure, in up to 26% of the patients who are undergoing dialysis for kidney disease. The gene transmission process is not completely understood but it is supposed to be polygenic mechanism and expresses itself in preferable environment suggesting gene-environment interaction. Genome wide-association (GWAS) studies have provided mixed results about gene expression.

Methodology: A case-control study methodology was implemented in this study. 1050 patients of diabetes mellitus type 2 (DMT2) were selected from Pakistan Institute of Medical Sciences PIMS, department of medicine. The cases were divided into two groups of early and advanced DKD (albuminuria 30-299mg/24h and \geq 300mg/24h respectively). There were 370 controls, 352 cases with early DKD and 328 cases with advanced DKD. DNA was isolated from peripheral blood and FRMD3 expression in kidney tissue samples was studied through kidney biopsy samples.

Results: Among all types of SNPs one was significantly associated with the particular disease and that was rs1888747, rs10868025 did not show any association on statistical analysis. The associations were observed in recessive models. To perform regression, the model was adjusted for diabetes type II duration, gender of the patient, BP readings and lipid file, the association still persisted. None of the other SNPs were found associated with this disease.

Conclusion: The FRMD3 protein expression negatively correlates with FRMD3 genes, so it is a good candidate gene for the development of albuminuria. Further exploration is suggested to determine the exact mechanisms for correlation.

INTRODUCTION

Diabetes mellitus is characterized by increased blood sugar levels consistently. The high amount of glucose in the body affects different organs like kidney causing diabetic kidney disease. Diabetic kidney disease (DKD) may cause renal failure, in up to 26% of the patients who are undergoing dialysis for kidney disease. (1–5) Diabetic kidney disease also

affects the cardiovascular tissues causing mortality and morbidity after cardiac surgery.(6,7) The development of diabetic kidney disease has genetic basis and there has been a lot of research on it recently.(8,9) The gene transmission process is not completely understood but it is supposed to be polygenic mechanism and expresses itself in preferable environment suggesting gene-environment interaction.(8–13) Genome wide-association (GWAS) studies have provided mixed results about gene expression. GWAS has identified 13 SNPs associated with DKD in two different populations having diabetes type I.(14)

The strongest association is of genes on FRMD3 locus. Other gene loci were also studied for associations. The populations affected with type I or type II diabetes mellitus may share a common gene loci related to DKD. (15–17)Our study will observe whether the SNPs that are associated with type I are also associated with type II diabetes or not. It will also evaluate the genes expressions from kidney biopsies and find the associations of SNPs with early and advanced stages of DKD.

METHODOLOGY

A case-control study methodology was implemented in this study. 1050 patients of diabetes mellitus type 2 (DMT2) were selected from Pakistan Institute of Medical Sciences PIMS, department of medicine. The cases were defined according to WHO definition of Diabetes mellitus; (18)patients with diabetes diagnosed at or above 35 years of age, no insulin administration for one year after diagnosis and no ketoacidosis. 1050 patients were further divide in to cases and controls. The controls were those patients who had no DKD and whose albumin was <30mg/24h. The cases were divided into two groups of early and advanced DKD (albuminuria 30-299mg/24h and \geq 300mg/24h respectively). There were 370 controls, 352 cases with early DKD and 328 cases with advanced DKD. The study protocol was presented to ethical committee of PIMS and executed after approval. Consent forms were signed by all the patients. The patients were evaluated physically and information was obtained about patient's age at diagnosis, drug treatment, smoking, history of other medical conditions and there treatment. The body mass index for each patient was calculated after measuring weight and height carefully. For cases with advanced DKD, the weight for dialysis dependent patients was measured by calculating mean of weights after dialysis in three consecutive sessions. Hypertension (BP \geq 140/90mmHg) records were obtained along with medication for BP. The blood samples were collected in fasting. Glucose oxidase method was used to determine fasting plasma glucose levels. Serum creatinine levels were measured through laboratory tests. Cholesterol and triglycerides were measured by enzyme methods. Immunoturbidimetry with intra and inter-assays was performed for urine analysis (urinary albumin excretion UAE).(19–22)

DNA isolation and genotyping

DNA was extracted by salt-out procedure from peripheral leukocytes. Our gene of interest was rs1888747 and rs10868025 both located on the same locus on 9q. Human Custom TaqMan Genotyping assays were used. Real time polymerase chain reaction RT-PCR was done in 96-well plates with primers and probes by using 5 microliters reaction solution and 2 nan-grams of DNA genome. The plates were placed on RT-PCR cycler on 95 degree Celsius and heated for 10 minutes. This was followed by 45 cycles on 95 degree for 15-20 seconds and then on 60 degree for one minute. The success rate was 95% with 0.01% calculated error. FRMD3 expression in kidney tissue sample. The DNA samples of patients with advanced DKD obtaining treatment for nephropathy during last one year were taken. Complete history of the patient was considered before DNA collection that included record of any previous diseases, record of nephrectomy, time since the diagnosis of diabetes mellitus and records of hypertension and smoking. mRNA expression using western blot (WB) and protein expression using immunohistochemistry (IHC) was performed for DNA samples obtained from kidney tissues. The kidney tissues was obtained after consent of ethical committee of hospital and written informed consent of the patient. Out of 90 samples collected, 45 samples were chosen for DNA genotyping others could not be used because the tissues were not enough.

Kidney biopsies were contained in phenol-isothiocyanate.(23) The RNA was extracted and centrifuged (12,000xg) with chloroform and isopropanol respectively. The precipitated RNA was then washed and suspended with ethanol (75%) and 10 – 50 microliter of diethylpyrocarbonate. The isolated RNA was then assessed on NANODROP

2000spectrophotometer.(24) The mean and standard deviation was $2.58 \pm 1.47 \mu\text{g}/250$ for the concentrated RNA obtained from kidney.

RT-PCR reverse transcription was done in two parts. The RNA was transcribed into cDNA and cDNA was amplified through reverse PCR transcription. The kit used was Super Script Vilo Master Mix Kit and the experiment was done in 7500 real time polymerase chain reaction system. $10 \mu\text{l}$ of SYBER Green solution was used in PCR reaction with $1 \mu\text{l}$ of gene specific primer (FRMD3), $7 \mu\text{l}$ of water and $1 \mu\text{l}$ cDNA. Three thermo-cycles were performed, first on 95 degree for ten minutes, then fifty cycles on 95 degree for 15 seconds each and last cycle on 95 degrees for one and half minutes. $\Delta\Delta\text{Cq}$ method was used to quantify FRMD3(25,26) cDNA and it was expressed to cyclophilin A in 90 kidney tissue samples, 74 of which were carrying G allele of rs1888747 SNP, and 16 with CC genotype. cDNA sample was diluted and amplification was done separately for FRMD3 and cyclophilin A. Equal amplification efficiencies were applied in all experiments (95-105%). $\Delta\Delta\text{Cq}$ method gives values for changes in gene expression with each fold. (27)

FRMD3 gene distribution was determined in forty seven samples due to technical limitations and inadequate sample. 39 of these samples carried allele for rs1888747SNP and 7 contained rs10868025.

To find the association between cases and controls first the cases and controls were kept into two different groups and then cases divided into further two groups with early and advanced diabetic kidney disease data were analyzed for best fit all frequencies of alleles of SNPs were calculated by counting genes and the distributions were verified using chi-square test. The distribution of genotype frequencies and alleles were compared using chi-square test. Mean and standard deviation values were used for continuous variables and medians were taken for the variables with skewed distributions. Categorical data were expressed in numbers of cases and percentage of individuals affected and ANOVA or one way analysis of variance. Chi-square test or student t-test were performed to evaluate the difference is clinical and laboratory representation of FRMD3 gene expression. Association magnitude was determined using odds ratio with 95% confidence interval (CI). Multinomial logistic regression analysis was used to determine the possibility of association of SNPs directly to kidney disease, co-variables were adjusted. Pearson chi-square value of correlation was used do valuate the correlation between variables. Multiple layered linear regression models were applied to FRMD3 gene as dependent variable and age and sex as independent variables. Result values less than 0.05 were considered significant. All analysis were performed as on SPSS

RESULTS

We evaluated 1050 patients of diabetes type two out of which 352 had diabetic kidney disease (DKD) of early stage and 328 had advanced diabetic kidney disease (DKD) and they were called cases. Other 370 patients were controls. Patients who had diabetic kidney disease (CKD) had bad profile in terms of blood pressure records and a bad lipid profile when compared to control group. Among all types of SNPs one was significantly associated with the particular disease and that was rs1888747, rs10868025 did not show any association on statistical analysis. The associations were observed in recessive models. To perform regression, the model wars adjusted for diabetes typeII duration, gender of the patient, BP readings and lipid file, the association still persisted. None of the other NSPs were found associated with this disease.

FRMD3 expression study

Since only rs1888747 was associated with diabetic kidney disease in this sample of subjects with diabetes type 2 and this SNP is located near FRMD gene we concluded that FRMD gene and protein expression in human kidney tissue is important to determine the prevalence of DKD. The mean age of 90 subjects for rs1888747 NSPs was 57.7 ± 14.2 . 57.8% were male, 50.4% had arterial HTN 22.8% were smokers and 18.9% had bad lipid profile.

Table 1 Genotype distribution of rs1888747

	Renal status		
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Genotype	Normal albuminuria N=370	Micro albuminuria N=352	Macro albuminuria N=328	P*	P**
CC	40 (10.18)	27 (7.7)	23 (7.01)	0.04	0.05
GC	152 (41.08)	148 (42.04)	146 (44.51)		
GG	178 (48.10)	177 (50.28)	159 (47.56)		
C allele	0.31	0.28	0.27		

The distribution is expressed as numbers and percentages
P* is obtained by all comparing all three groups
P** is obtained by comparing micro and macro-albuminuria

Table 2 Genotype distribution in cases and controls

Polymorphism	Controls (n = 370)	Cases (n = 680)	P	Dominant: OR (95 % CI)/P	Recessive: OR (95 % CI)/P	Additive: OR (95 % CI)/P
CC	10.18 (40)	50 (7.4)	0.037	0.81 (0.71– 1.05)/0.032	0.59 (0.40– 0.93)/0.028	0.59 (0.37– 0.89)/0.029
CG	41.08 (152)	294 (43.2)				
GG	48.10 (178)	336 (49.41)				

The distribution is expressed as frequencies and percentages

Since rs1888747 was the only SNP associated significantly with DKD (CC/CG/GG = cases vs. controls = 7.4%/50 vs 10.18%/40 P = 0.037). Minor allele frequencies were 0.31 and 0.55 in controls and cases (microalbuminuric + macroalbuminuric/ESRD), respectively (P = 0.06). The strongest association was observed assuming recessive (CC vs. CG/GG, OR: 0.59, 95 % CI 0.40–0.93; P = 0.028) and additive (CC vs. GG, OR: 0.59, 95 % CI 0.37–0.89; P = 0.029) models. This association persisted after controlling for T2DM duration, gender, systolic BP, and triglycerides profile. The other SNPs are not associated with DKD including rs10868025. No other allele was precipitated or showed positive results so it is assumed that they were not expressed in DKD patients.

DISCUSSION

In our study, it was found that rs1888747 is positively associated with DKD in type 2 diabetes mellitus; it is associated with type1 diabetes as stated in previous studies.(15) The DKD patients were further divided into two groups to explore the reasons for development of DKD. The CC genotype is dominant in microalbuminuria and shows protective behavior. GWAS has explored many unknown genes and alleles associated with DKD but these studies were not replicated for different populations. (13)(28,29)The gaps in the studies have led to lesser evidences on the causation of the condition and association of the genes. rs1888747 SNP is associated with development of DKD but the populations studies in different ethnic regions have produced confusing results. (30–34)

GWAS has been studying genes on different locations, there have been advances in genetic engineering and various genotypes are studied over the years but the exact pathogenesis of DKD remains elusive. Gene-environment interactions, phenotypes, different characteristics of populations. This study also aimed at evaluating risk related to rs1888747 alleles by determining the influence on FRMD3 gene expression. FRMD3 is located on 9q21.32. The FRMD3 protein expression negatively correlates with FRMD3 genes, so it is a good candidate gene for the development of albuminuria. Further exploration is suggested to determine the exact mechanisms for correlation.

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