

# MICROBIAL EVALUATION AND ANTIBIOTIC SUSCEPTIBILITY OF PATHOGENIC BACTERIA ISOLATED FROM “PLA-SOM” AND “PLA-CHOM”, THAI RICE-FERMENTED FISH PRODUCTS

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## Abstract

Pla-som and pla-chom are Thai traditional fermented fish, which is produced from fish, sugar, salt and roasted rice and is fermented with natural lactic acid bacteria, therefore pathogenic bacteria isolation from these traditional products are sparsely reported. This study was aimed to evaluate contamination of pathogenic bacteria and their antibiotic susceptibility from pla-som (n = 15) and pla-chom (n = 15). According to colony, Gram staining and biochemical characteristics, pathogenic bacteria were isolated and identified; tested for antibiotic susceptibility. Pathogenic bacteria isolate from pla-som (n = 32) were higher than pla-chom (n = 12). Suspected pathogenic bacteria were *Staphylococcus aureus* and *Escherichia coli* isolated from both of rice-fermented fishes while *Salmonella typhimurium*, *Listeria monocytogenes* and *K. pneumoniae* were commonly contaminated in pla-som. Salt-tolerant pathogens including *Vibrio* sp. and *Aeromonas hydrophila* were also identified. Fifty and twenty of pathogenic bacteria were isolated from pla-som and pla-chom, which were determined on susceptibility with fifteen antibiotic discs. Some of pathogenic bacteria were resistant to three or more antibiotics and defined as multidrug resistance (MDR) bacteria. Various pathogenic bacteria including *S. aureus*, *E. coli*, *K. pneumoniae*, *S. typhimurium*, *A. hydrophila*, *Vibrio* sp. and *Proteus* sp., were antibiotic resistance. *Vibrio* sp. and *A. hydrophila* were MDR bacteria with five or more antibiotic resistances. The finding is notified on awareness of recipe in consumers and regarding on food safety of traditional fermentation process in local manufacturers.

**Keywords:** pla-som, multidrug resistance (MDR), pathogenic bacteria, Thai fermented fish.

## INTRODUCTION

Fermentation is common food preserving method, which is inhibit of spoilage microbial growth by organic acids and by-product metabolites. As a result, these products generally have a longer shelf life than their original food [1, 2]. Fermented food is providing health promotion including lowering of blood cholesterol, immunomodulation, etc. However, food-borne illness from fermented food consumption had reported, which is due to contamination of bacteria and its toxins [3].

Particularly, food-borne illness is occurred in food products from local household fermentation [4]. Pla-som is Thai traditional fermented fish, which is produced from fish, sugar, salt and roasted rice (sometimes contained with spices) and is fermented with natural microbial flora.

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Ingredient and process of pla-chom is similar to pla-som, however there is use tiny fishes as raw material, while pla-som is processed from large flesh or whole of fish. Amount of lactic acid bacteria are preserving pla-som, which is depended on salt concentration [5]. Recently, of traditional Thai fermented foods are lack of commercialize starter culture of lactic acid bacteria and majority of fermented foods are produced from small scale production [6]. According to the Thai Agricultural Standard, Thai fermented fish should meet in criteria of requirements including physical characteristics, salt content, food additives, and contaminants i.e., toxic metals and microbials. Traditional fermentation processes i.e., raw ingredient preparation, personnel hygiene and practices, food packaging which is risk to contaminate with various bacteria including pathogenic bacteria [3, 7]. Pathogenic bacteria including *Bacillus cereus*, *Clostridium perfringens*, *Escherichia coli*, *Salmonella* spp., and *Staphylococcus aureus* are restricted bacterial contaminants according to the Thai Agricultural Standard [8, 9]. Isolation of pathogenic bacteria from pla-som and pla-chom, Thai rice-fermented fish products are sparely reported when compare with plara, Thai fermented fish without rice contained [8, 9]. The antibiotic resistance of isolated bacteria is also interesting to determine in this study. Because of their can affect public health due to transmission or cross-contamination of foodborne diseases, which are depended on fermentation process, personnel hygiene and device-related contaminants [10, 11]. Hence, we were concerning on bacterial contamination isolated from this Thai rice-fermented fish products. Isolated pathogenic bacteria were further determined antibiotic susceptibility. The finding was provided information and the guideline for local food manufacturers for controlling of fermentation process, which is meet to food quality. In addition, the finding will report and notify on consumers and public health policy on controlling of foodborne diseases.

## MATERIALS AND METHODS

### Sample Collection and Preparation

Pla-som (n = 15) and pla-chom (n = 15) were purchased from six markets located in Bangkok, Thailand. Each fermented fish was weighed (25 g), suspended in 225 ml of 0.1% peptone water (Merck, Darmstadt, Germany) for enrichment (sample-to-broth ratio, 1:10) and inoculated in selective media (Himedia, India) including McConkey agar, xylose lysine deoxycholate (XLD) agar and eosin-methylene blue (EMB) agar at 37 °C for 24 hrs. Isolated colony was sub-cultured in tryptic soy agar (TSA) agar and incubated similar to selective media for biochemical and antibiotic susceptibility tests.

### Bacterial Colony Isolation and Biochemical Identification

Bacterial colony isolate was characterized according by size, shape, convex, edge, color, and morphology. Each isolated

colony was stained and identified by Gram □s staining and biochemical tests, respectively. Biochemical tests were included catalase, oxidase, string test, citrate, methyl red, Voges-Proskuer test, oxidation-fermentation, lysine, indole, motility, bile esculin, triple sugar iron (TSI), which were identified pathogenic Gram □s positive bacteria, Gram □s negative Enterobacteria and non-fermentative bacilli (NFB). Thiosulfate citrate bile salt sucrose agar (TCBS) and 0%, 3%, 6%, 8% and 10% NaCl in nutrient broth were used to identified *Vibrio* spp.

Colony, Gram □s staining and biochemical characteristics of bacterial isolates were interpreted and identified according by Bergey□s Manual of Systematic Bacteriology and Clinical and Laboratory Standards Institute (CLSI) [12, 13].

### Antibiotic Susceptibility Test

Pathogenic bacteria were evaluated on antimicrobial susceptibility of by agar disc diffusion (Kirby-Bauer) method. Each isolated bacterial colony was sub-cultured in TSA agar, and was inoculated into 4 mL of sterile nutrient broth. Bacterial suspension was adjusted its density with 0.5 McFarland as standard. Then, bacterial suspension was spread over entirely surface of Muller Hinton (MHA) plate, which was dried by standing at room temperature. Fifteen of antibiotic discs (Difco, USA) used in susceptibility test were included amoxicillin/clavulanic acid (30 □g, 20/10 □g), ampicillin (10 □g), amikacin (30 □g), cefoxitin (30 □g), ceftazidime (30 □g), ceftriaxone (30 □g), cefuroxime (30 □g), chloramphenicol (30 □g), ciprofloxacin (5 □g), gentamicin (10 □g), imipenem (10 □g), meropenem (10 □g), piperacillin (30 □g) tetracycline (30 □g), trimethoprim/sulfamethoxazole (25 □g). Inhibition zone was clearing zone surrounded antibiotic disc and measured as diameter (mm). Each inhibition zone value was compared with CLSI reference and interpreted as susceptible (S), intermediate (I) and resistance (R) bacteria against tested antibiotic disc. For this study, intermediate antibiotic susceptibility bacteria were considered as antibiotic resistance and multi-drug resistance (MDR) bacteria were defined as isolated bacteria is resisted three or more different antibiotic discs [14].

## RESULTS AND DISCUSSION

### Bacterial Identification and Distribution

Examples of bacterial colonies in each selective media were represented in Fig. 1. Selective media were isolated mainly members of Enterobacteriaceae according by sugar fermentation pattern, color of pH indicators contained in each media, and also pigment formation. According from colony, Gram □s staining and biochemical characteristics, pathogenic bacteria were isolated and identified. Number of pathogenic bacteria isolates from pla-som (n = 32) were higher rather than pla-chom (n = 12), which were unrelated

to markets and type of pathogenic bacteria. Suspected pathogenic bacteria were *S. aureus* and *E. coli* isolated from both of rice-fermented fishes while *S. typhimurium*, *L. monocytogenes* and *K. pneumoniae* were commonly contaminated in pla-som. We were also found some of *Vibrio* sp. and *Aeromonas hydrophila* isolates from pla-som and pla-chom (Table 1).

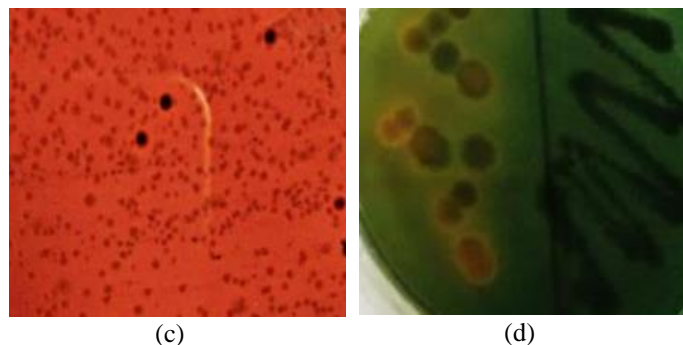
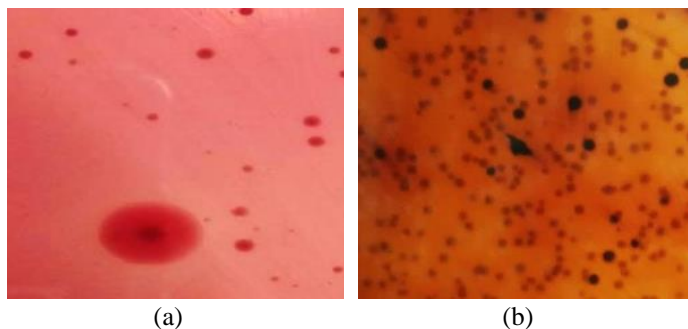


Fig. 1: Bacterial colonies were represented in selective media: (a) McConkey agar, (b) XLD agar, (c) EMB agar (d) TCBS agar

Table 1: Distribution of pathogenic bacteria isolates from Thai rice-fermented fish

Identification/ Fermented fish	Pla-som (n = 15)			Pla-chom (n = 15)		
	M1	M2	M3	M4	M5	M6
<i>Staphylococcus aureus</i>	4	1	2	-	-	1
<i>Bacillus cereus</i>	-	-	1	-	1	-
<i>Listeria monocytogenes</i>	1	2	-	-	-	-
<i>Escherichia coli</i>	3	5	3	-	2	-
<i>Salmonella typhimurium</i>	2	-	1	-	1	-
<i>Klebsiella pneumoniae</i>	-	1	3	1	-	-
<i>Proteus mirabilis</i>	-	-	1	-	2	-
<i>Proteus vulgaris</i>	2	-	1	-	1	-
<i>Enterobacter aerogenes</i>	1	-	-	-	-	-
<i>Vibrio cholerae</i>	-	-	-	1	-	2
<i>Vibrio parahaemolyticus</i>	1	-	-	-	-	1
<i>Vibrio mimicus</i>	1	-	-	-	-	-
<i>Vibrio vulnificus</i>	1	-	-	-	2	-
<i>Vibrio harveyi</i>	2	-	-	-	-	1
<i>Aeromonas hydrophila</i>	1	1	-	1	-	-
<i>Pseudomonas</i> sp.	1	-	-	-	-	-
Total for each market (M)	20	10	12	3	5	4
Total for fermented fish	32			12		
<i>Lactobacillus</i> sp.	3	5	1	-	3	-

Hence, this salt-tolerate pathogens can grow, whereas this product was made from freshwater fish and its salinity was came from variety of salt ingredient. *Lactobacillus* sp. was contained only in some of products. There was implied that organic acid and preservative metabolites in this product were produced from *Lactobacillus* sp. and other lactic acid bacteria (LAB). Pla-som and sour fish products in South-East Asia region are contain LAB including *Lactobacillus*, *Lactococcus*, *Pediococcus*, *Streptococcus*, *Enterococcus* and *Weissella* sp. [15-17]. There can inhibit foodborne pathogens and food spoilage microorganisms in pla-som by bacteriocin production i.e., *S. salivarius* and *E. faecalis* can inhibit *E. coli*, *S. aureus* and *Salmonella* sp. [15]; *W. cibaria* 110 can inhibit some of Gram-positive bacteria by *Weissellicin* 110 (bacteriocin) production [18]. Pathogenic *Vibrio* sp., *S. typhimurium* and *L. monocytogenes* were

contaminated in pla-som that may out of standard according by Food and Agriculture Organization (FAO). *S. aureus* and *E. coli* were also contaminated in some of rice-fermented fish products, which were concerned on food safety. Therefore, there was lack of quantitative data, which was unable to determine food safety of *S. aureus* and *E. coli* contamination in this study [8, 9, 19]. Contamination of *Vibrio* and *Salmonella* sp. had also reported in other Thai fermented fish contained garlic or “som-fak” and inhibition of bacterial contamination is depended on amount of lactic acid, salt and garlic [20]. Food quality is not concerning only pathogens and its toxins that cause illness, there is also consider on preferable shelf-life [20]. In case of this study, Thai rice-fermented fish was still desirable characteristics such as flavor, texture and shelf-life. Thus, pathogenic contamination during and after fish fermentation process is

one of factor that need to concern [19, 20, 21]. Moreover, acceptability of local fermented food is also important in different cuisine culture. In Thailand and South East Asian countries, fermented food products are valuable by some culture, therefore there are looked disgusting by others. Cuisine culture in North-East of Thailand and nearby area is mostly consumed fermented fish as dietary protein source, however this preserved food has strong odor and flavor as spoilage-like [22, 23].

#### Antibiotic Susceptibility of Pathogenic Bacteria Isolates

Example of susceptibility test in pathogenic bacterial isolates was shown in Fig. 2. Fifty and twenty of pathogenic bacteria were isolated from pla-som and pla-chom, which were determined on susceptibility with fifteen antibiotic discs. Some of pathogenic bacteria were resisted to three or more antibiotics and defined as multidrug resistance (MDR) bacteria. MDR bacteria isolates from pla-som were *E. coli* (n = 5, 45%), *S. aureus* (n = 4, 57%), *S. typhimurium* (n = 2, 67%), *P. vulgaris* (n = 2, 67%), *V. mimicus* (n = 1, 100%),

*V. vulnificus* (n = 1, 100%), *V. harveyi* (n = 1, 50%), *A. hydrophila* (n = 1, 50%) and *K. pneumoniae* (n = 1, 33%). MDR bacteria isolates from pla-chom were *V. vulnificus* (n = 2, 100%), *A. hydrophila* (n = 1, 100%), *S. typhimurium* (n = 1, 100%), *P. mirabilis* (n = 1, 50%) and *E. coli* (n = 1, 50%). Some of isolates including *V. mimicus*, *V. vulnificus* and *K. pneumoniae* isolated from pla-som, and *V. vulnificus*, *A. hydrophila* and *E. coli* were rarely occurred MDR bacteria, however there were against resisted five or more antibiotics (Table 2).

In Thailand, pathogenic bacteria contamination in food is common causes of gastro-intestinal diseases. Antibiotic resistance in foodborne microorganisms is worldwide public health concern [24]. Overuse and misuse of antibiotics in livestock farming are remain in environments and there is transfer to water resources and aquaculture [25]. Various microorganisms in fermented foods may act as carriers of antibiotic resistance genes, which are transferred through in food chain between environments, food and human.

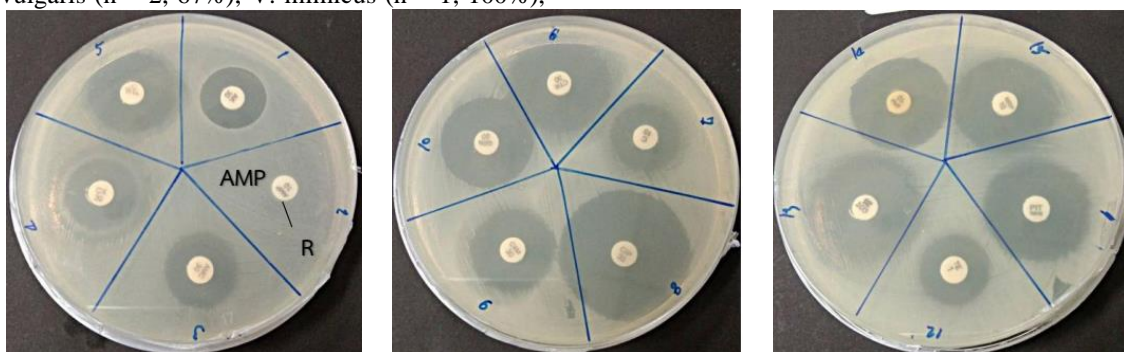


Fig. 2: Bacterial culture was susceptible to fourteen of antibiotic discs and represented as inhibition (clear) zone (mm). Inhibition zone surrounded ampicillin (AMP) disc was narrower than standard range and interpreted as ampicillin resistance (R).

Table 2: Antibiotic resistance patterns of pathogenic bacteria isolated from pla-som and pla-chom

Sample/ Isolates	Antibiotic resistance pattern*	Resistance (%)	MDR (%)
<b>Pla-som</b>			
<i>E. coli</i> (n = 11)	AMP-TE-COT; CX-C-TE; AMP-C-TE; AMP-C-TE; AMP-CXM; AMP-CX; AMP-C; C-CXM; AMP	10 (91%)	5 (45%)
<i>S. aureus</i> (n = 7)	AMP-CX-C-CXM-TE; AMP-CX-TE; AMP-PI-TE; AMP-PI-TE; CIP-TE; AMP; AMP	7 (100%)	4 (57%)
<i>K. pneumoniae</i> (n = 4)	AK-AMP-C-GEN-PI; AMP-CXM; AMP-CX	3 (75%)	1 (33%)
<i>S. typhimurium</i> (n = 3)	AMP-CXM-TE; C-TE-COT; TE	3 (100%)	2 (67%)
<i>P. vulgaris</i> (n = 3)	AMP-CX-CXM-TE-IPM; C-CXM-TE; AMP	3 (100%)	2 (67%)
<i>L. monocytogenes</i> (n = 3)	GEN	1 (33%)	-
<i>A. hydrophila</i> (n = 2)	AMP-TE-IPM; AMP-TE	2 (100%)	1 (50%)
<i>V. harveyi</i> (n = 2)	AMP-CX-TE	1 (50%)	1 (50%)
<i>V. mimicus</i> (n = 1)	AMP-CX-C-CXM-TE	1 (100%)	1 (100%)
<i>V. vulnificus</i> (n = 1)	AMP-CTR-C-CXM-TE	1 (100%)	1 (100%)
<i>E. aerogenes</i> (n = 1)	AMP-C	1 (100%)	-
<i>Pseudomonas sp.</i> (n = 1)	AMP	1 (100%)	-
<b>Pla-chom</b>			
<i>V. cholerae</i> (n = 3)	AMP	1 (33%)	-
<i>V. vulnificus</i> (n = 2)	AMP-AMC-C-CXM-TE-COT-IMP; AMP-C-CXM-COT-IMP	2 (100%)	2 (100%)

<i>P. mirabilis</i> (n = 2)	AMP-TE-COT; AMP-TE	1 (100%)	1 (50%)
<i>E. coli</i> (n = 2)	AK-AMP-CX-CAZ-CTR-CXM-TE; TE	1 (100%)	1 (50%)
<i>S. typhimurium</i> (n = 1)	AMP-C-TE	1 (100%)	1 (100%)
<i>V. parahaemolyticus</i> (n = 1)	AMP	1 (100%)	-
<i>V. harveyi</i> (n = 1)	CX	1 (100%)	-
<i>A. hydrophila</i> (n = 1)	AK-AMP-CAZ-PI-TE-COT	1 (100%)	1 (100%)
<i>S. aureus</i> (n = 1)	AMP	1 (100%)	-
<i>K. pneumoniae</i> (n = 1)	CX	1 (100%)	-

\*Pattern of antibiotic drug resistance for each isolate and MDR was defined as bacterial resistant against three or more antibiotics

AK = amikacin; AMP = ampicillin; AMC = amoxiclav; CX = cefoxitin; CAZ = ceftazidime; CTR = ceftriaxone; C = chloramphenicol; CIP = ciprofloxacin; CXM = cefuroxime; GEN = gentamicin; PI = piperacillin; TE = tetracycline; COT = co-trimoxazole; IPM = imipenem; MR = meropenem; MDR = multidrug – resistant bacteria

As our result, bacterial isolated from Thai fermented fish especially from pla-som were MDR characteristics. Thus, antibiotic resistance of pathogenic bacteria may become concerning in food safety of Thai fermented fish particularly from traditional process. In addition, lactic acid bacteria in Thai fermented fish products are also considered on genetic transferring of antibiotic resistances [26, 27, 28]. Further study is need to conduct on fermentation parameters of Thai fermented fish i.e., pH, salinity and lactic acid bacteria contained. Beta-lactamase genes and other common antibiotic resistance genes of MDR bacteria in Thai fermented fish products are also need to determined.

## CONCLUSION

Pathogenic bacteria and their antibiotics resistance were occurred in Thai rice-fermented fish, namely pla-som and pla-chom, which may risk of foodborne illness. Simultaneous antibiotic resistances of various pathogenic bacteria including *S. aureus*, *E. coli*, *K. pneumoniae*, *S. typhimurium*, *A. hydrophila*, *Vibrio* sp. and *Proteus* sp., were represented in this study. *Vibrio* sp. and *A. hydrophila* were rarely isolated MDR bacteria, therefore there were resisted against five or more antibiotics. We were concluded that these fermented fish products may a reservoir of antimicrobial resistance/virulence factors and can transfer through food chain by genetical transmission. The finding is notified on awareness of recipe in consumers and regarding on food safety of traditional fermentation process in local manufacturers.

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## REFERENCES

- Steinkraus KH. Industrialization of indigenous fermented foods. 2nd ed. New York: CRC Press; 2004.
- Adams M, Mitchell, R. Fermentation and pathogen control: a risk assessment approach. *Int J Food Microbiol* 2002;79(2):75e83.
- Nout MJR. Fermented foods and food safety. *Food Res Int* 1994;27(3): 291e298.
- Nout MJR, Motarjemi Y. Assessment of fermentation as a household technology for improving food safety: a joint FAO/WHO workshop. *Food Control* 1997;8(6):221e226.
- Østergaard A, Embarek PKB, Wedel-Neergaard C, Huss HH, Gram L. Characterization of anti-listerial lactic acid bacteria isolated from Thai fermented fish products. *Food Microbiology* 1998;15(2):223e233.
- Valyasevi R, Rolle RS. An overview of small-scale food fermentation technologies in developing countries with special reference to Thailand: scope for their improvement. *Int J Food Microbiol* 2002;75(3):231e239.
- Capozzi V, Fragasso M, Romaniello R, Berbegal C, Russo P, Spano G. Spontaneous food fermentations and potential risks for human health. *Fermentation* J 2017;3(4):49-51.
- Sangjindavong M, Chuapoe huk P, Runglerdkriangkrai J, Klaypradit W, Vareevanich D. Fermented fish product (pla-ra) from marine fish and preservation. *Kasertart J* 2008; 42:129-136.
- Rattanasuk S, Boonbao J, Sankumpa, Surasilp T. Foodborne pathogens in fermented fish purchased in Selaphum, Roi Et. *Intl Conf Sci Tech* 2015;1(3):178-181.
- Wirtanen G, Salo S. Microbial contaminants and contamination routes in the food industry. Finland: Julkaisija-Utgivare Publishing; 2007.
- Todd EC, Greig JD, Bartleson CA, Michaels BS. Outbreaks where food workers have been implicated in the spread of foodborne disease: transmission and survival of pathogens in the food processing and preparation environment. *J Food Protection*. 2009; 72:202-219.
- Brenner DJ, Krieg NR, Staley JT, Garrity GM. *Bergey’s Manual of Systematic Bacteriology*. Parts A, B and C. 2nd ed., Vol. 2. New York: Springer-Verlag; 2005.
- Clinical and Laboratory Standards Institute (CLSI). *Quality Assurance for Commercially Prepared Microbiological Culture Media*. CLSI; 2016.p. M22.
- Wayne P. Clinical and laboratory standards institute. *Performance standards for antimicrobial susceptibility testing*, 2011.
- Hwanhlem N, Buradaleng S, Wattanachant S, Benjakul S, Tani A, Maneerat S. Isolation and screening of lactic acid bacteria from Thai traditional fermented fish (Plasom) and production of Plasom from selected strains. *Food Control* 2011;22(3-4), 401-407.
- Kopermsub P, Yunchalard S. Identification of lactic acid bacteria associated with the production of plaa-som, a traditional fermented fish product of Thailand. *Int J Food Microbiol* 2010;138(3):200-

204.

17. Marui J, Boulom S, Panthavee W, Momma M, Kusumoto K-I, Nakahara K, et al. Culture-independent analysis of the bacterial community during fermentation of pa-som, a traditional fermented fish product in Laos. *Fish Sci* 2014;80(5) :1109-1115.
18. Sriornual S, Yanagida F, Lin LH, Hsiao KN, Chen YS. Weissellicin 110, a newly discovered bacteriocin from *Weissella cibaria* 110, isolated from pla-som, a fermented fish product from Thailand. *Appl Environ Microbiol* 2007;73(7):2247-2250.
19. FAO. Biotechnologies for Agricultural Development: Proceedings of the FAO international technical conference on “Agricultural Biotechnologies in Developing Countries: options and opportunities in crops, forestry, livestock, fisheries and agro-industry to face the challenges of food insecurity and climate change” (ABDC-10), Rome: FAO; 2011.
20. Bernbom N, Ng YY, Paludan-Müller C, Gram L. Survival and growth of *Salmonella* and *Vibrio* in som-fak, a Thai low-salt garlic containing fermented fish product. *Int J Food Microbiol* 2009;134(3):223-229.
21. Adams M, Mitchell R. Fermentation and pathogen control: a risk assessment approach. *Int J Food Microbiol* 2002;79(2):75e83.
22. Rozin P, Haidt J, McCauley CR. Disgust. In M. Lewis, J. M. Haviland-Jones, & L. F. Barrett (Eds.). *Handbook of emotions* New York: Guilford Press; 2008, p. 757-776.
23. Beddows CG. Fermented fish and fish products. In J. B. B. Wood (Ed.). *Microbiology of fermented foods*. Glasgow, UK: Blackie Academic and Professional; 1998, p. 416-440.
24. Abriouel H, Knapp CW, Gálvez A, Benomar N. Antibiotic resistance profile of microbes from traditional fermented foods. In J. Frias, C. Martínez-Villaluenga, & E. Peñas (Eds.). *Fermented foods in health and disease prevention*. London, UK: Academic Press; 2017, p. 675-704.
25. Van Boeckel TP, Brower C, Gilbert M, Grenfell BT, Levin SA, Robinson TP, et al. Global trends in antimicrobial use in food animals. *Proc Natl Acad Sci USA* 2015;112(18):5649-5654.
26. Sornplang P, Leelavatch V, Sukon P, Yowarach S. Antibiotic resistance of lactic acid bacteria isolated from a fermented fish product pla-chom. *Res J Microbiol* 2011;6(12):898-903.
27. Li SW, Chen YS, Lee YS, Yang CH, Sriornual S, Wu HC, et al. Comparative genomic analysis of bacteriocin-producing *Weissella cibaria* 110. *Appl Microbiol Biotechnol*. 2017 Feb;101(3):1227-1237.
28. Ishibashi N, Himeno K, Masuda Y, Perez RH, Iwatani S, Zendo T, et al. Gene cluster responsible for secretion of and immunity to multiple bacteriocins, the NKR-5-3 enterocins. *Appl Environ Microbiol* 2014;80(21):6647-6655.