

STUDY EFFECT OF TAMOXIFEN ON CELL CYCLE PHASE DISTRIBUTION, IN BREAST CANCER CELLS

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Abstract

Hormone therapy also known as estrogen suppression therapy is the first targeted therapy and is frequently used as a second therapeutic to lower the risk of cancer returning after surgery or spread. Tamoxifen is a drug that is commonly used to treat breast cancer by mediating estrogen receptor signaling pathways. Breast cancer is becoming a spread disease that needs a specific therapy. The main focus of this study is the anti-cancer drug Tamoxifen (TAM) and its effect on cell cycle (Culture) of breast cancer cell line measurement of cellular DNA content and the analysis of the cell cycle can be performed by flow cytometry. This study includes the determination of the effects of tamoxifen on MCF-7i on breast cancer cell line and investigate of the mechanism by which it is affected on the cell cycle phases using Flow cytometric cell cycle analysis assay. As well study the Flow cytometric cell cycle analysis of tamoxifen-treated cultures showed an increase in the proportion of cells in the G1 phase of the cell cycle.

Keywords: Breast cancer, Tamoxifen, Flow cytometry, Cell cycle.

INTRODUCTION

Cancer is a disease described by uncontrolled development of the cells. It can develop within any organic site in the body but the most common types include lung, breast, prostate and colorectal cancers (1). The American Cancer Society calculates the number of new cancer cases and fatalities in the United States each year, In 2022, approximately 1,918,030 new cancer cases and 609,360 cancer deaths (2). Breast cancer is the most common cancer in women worldwide. About 70-75% of breast tumors are estrogen receptor (ER) and/or progesterone receptor (PR)-positive (3). It is a heterogeneous disease, which comprises of many biologically different entities with distinct pathological features and clinical implications (4,5). Breast cancer appears in the form of tumor when there is uncontrollable proliferation of breast cells (6).

It can be classified into non-invasive breast cancer (Ductal carcinoma in situ and Lobular carcinoma in situ) and invasive breast cancer (invasive ductal carcinoma, invasive lobular carcinoma, Paget's disease of the nipple and inflammatory breast cancer) (7). The treatment of breast cancer are basically depends on tumor staging. There are different types of treatments that are used separately or in combinations, such as surgery (removal of the cancer tissue), radiation therapy, and systemic treatments. Breast cancer is a heterogeneous disease and treatment methods depend on different clinical criteria e.g. age, type of cancer, size, stage of detection and metastasis (8).

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Hormonal therapy is used usually in treating the patients with positive tumor after the completion of chemotherapy. One of the most important anticancer drug is Tamoxifen (TAM) used to treatment of breast cancer. Chemically known as (z)-2-[4-(1, 2-Diphenyl-1-butenyl) phenoxy-N,N-Dimethylethanamine (9). It is a selective estrogen receptor modulator (SERM) (10). Tamoxifen inhibits breast cancer cell growth and induces apoptosis. Apoptosis, or programmed cell death, is one of the topics that cell biologists are most interested in. It's a well-ordered and well-organized cellular process that happens in both normal and pathological settings and entails a sequence of biochemical processes that lead to a certain cell shape and death (11). The different stages of apoptotic cell death, started by cellular shrinkage and chromatin condensation, associated with formation of membrane blebbing, organelles and nucleus fragment and the blebs begin formation of apoptotic bodies which are finally phagocytosed by macrophages or neighboring cells by endocytosis/phagocytosis. Apoptosis plays a role in preventing cancer. If a cell is unable to undergo apoptosis due to mutation or biochemical inhibition, it can continue dividing and developed into a tumor (12). The most common and commonly used treatment for breast cancer is tamoxifen, which causes tumor stabilization in 50% of all patients with metastatic breast cancer who were previously untreated. Of all breast tumor types, 75% are ER positive. (13). Breast cancer cells may undergo apoptosis when treated to TAM via the membrane receptor pathway. The purpose of this study was to use flow cytometry cell culture to investigate about how TAM affected MCF 7i (ER+) cells'. Flow cytometry (FCM), a quantitative single cell analysis technique based on impedance or laser technology for quantitative single cell analysis that is used in cell sorting, cell counting, and biomarker identification. It provides the multi-parameter examination of single cells at the same time (14). The main clinical use of flow cytometry is the screening of hematologic malignancy, but it also has a broad range of additional uses, including counting reticulocytes and analyzing cell function and the cell cycle. G1 phase refers to the replication (Gap 1). During the S (Synthesis) phase, DNA replication takes place. Before mitosis, there is another interval (G2 phase, Gap 2) after DNA synthesis (M). Mature neurons and other cells that no longer divide are considered to have entered the G0 phase of the cell cycle (15). Propidium Iodide-A histogram with markers of G0/iG1, S, and G2/iM phases to find cell cycle phases, and markers of sub G0 /G1 and > (16). These *in vitro* study showed the effect of (TAM) on MCF-7 cells.

Materials and Methods

Propidium iodide was used as a DNA stain in the cell cycle analysis using the BD Cycletest™ Plus kit (BD Biosciences, San Diego, CA, United States). According to the manufacturer's instructions, flow cytometry was used

to assess the impact of Tamoxifen therapy on the cell cycle. Briefly, density-gradient centrifugation was used to separate MCF-7 cells. After centrifuging the cell suspension for five minutes at 300g at room temperature (25°C) while adding 200 L of fetal bovine serum in total, the entire tube of cell suspension was transferred into a 15 ml centrifuge tube. To prevent disrupting the particle, the supernatant was aspirated, leaving around 50 L of residual fluid in the tube. The cells were to be gently vortexed at low speed with 100 mL of buffer solution before being centrifuged for 5 minutes at 300 g at room temperature (25 °C). The supernatant was aspirated and leaving about 50 µL of residual fluid in the tube to prevent disturbing the pellet. The 1 ml of buffer solution was used to resuspend the cell pellet. A hemacytometer was used to count the cells after one additional centrifugation and resuspension of the sample. Buffer solution was used to adjust the concentration to 1.0×10^6 cells/mL. To achieve concentration to 1.0×10^6 cells/mL, Buffer solution was employed. This needs to generate enough cells for a test sample. Carefully glugging the whole supernatant, we next added 250 L of Solution A (trypsin buffer), and incubated for 10 minutes at 25°C. Each tube received 200 L of Solution B (trypsin inhibitor and RNase buffer), which was followed by 200 L of cold Solution C (8°C), which was incubated for 10 minutes at room temperature (PI stain solution). Distribution of the cell cycle nuclear DNA was determined by using flow cytometry (BD Biosciences). The proportion of cells in G1, S and G2 phases were examined by using Divani software (BD Biosciences).

Statistical analysis

Graph Pad Prism version 7 was used to analyze the data and represent it as a mean and standard deviation.

Results

The distribution of cell cycle was analyzed after treating the cells with tamoxifen at different concentrations. 6.25, 12.5 and 25 µg/mL that shown in Figure, (1), The histogram in figure A showing MCF-cell distribution at different phase, G1, S and G2/M, using, treatments control. In figure B Shows the all data and the mean of it. When using (6.25, µg/mL) of the drug, the G1 % \pm SD MCF-7 was equal to (57.2 \pm 0.566), in S was equal to (22.55 \pm 0.636) and in G2 (18.1 \pm 0.283). In (12.5, µg/mL) the \pm SD of G1, S, and G2 phases was equal (62.95 \pm 0.636), (20.65 \pm 0.495) and (14.75 \pm 0.354) respectively. The final concentration of tamoxifen drug that using was equal (25 µg/mL) and the \pm SD of data analysis in G1, S, and G2 shows (58.85 \pm 0.354), (19.85 \pm 0.212) and (19.35 \pm 0.212) respectively, all these data shown in table (1).

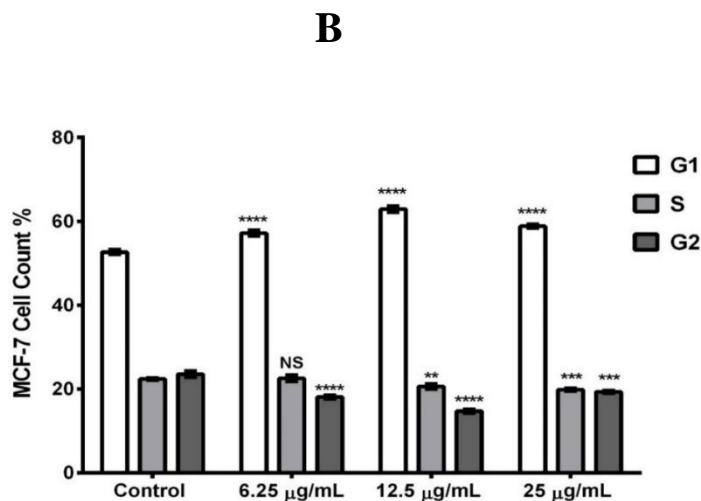
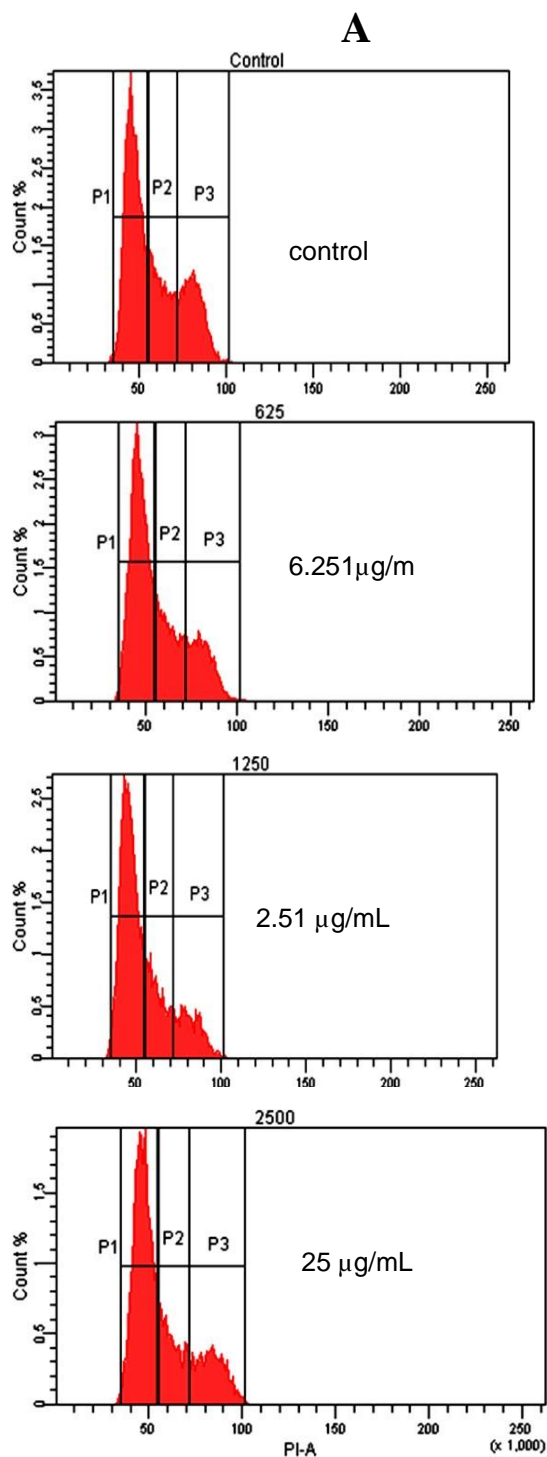


Figure 3-5 the, effect, of tamoxifen, on MCF-7 cells, cycle phase, distribution,]. (A) Flow cytometry histogram, showing MCF-cell, distribution, at different, cell cycle phase, G1, S and G2/M, using treatments, control, 6.25i, 12.5 and 25µg/mL of tamoxifen,. (B) Mean

Table 1: Tamoxifen effect on cell cycle in MCF-7 cells compering with control

Concentration µg/ml	G1 % ±SD MCF-7	S % ±SD MCF-7	G2 % ±SD MCF-7
Control	52.7 ±0.424	22.4 ±0.141	23.55 ±0.636
6.25	57.2 ±0.566	22.55 ± 0.636	18.1 ±0.283
12.5	62.95 ±0.636	20.65 ±0.495	14.75 ±0.354
25	58.85 ±0.354	19.85 ±0.212	19.35 ±0.212

Discussion

Cell cycle phase distribution of MCF-7 cells study was performed to determine whether or not tamoxifen suppression, of MCF-7 cell viability was related to cell cycle arrest (14). Flow cytometry was used to detect the DNA content after cell staining with propidium iodide. A and B showed that at G1 phase the percentage of cells was significantly increased by increasing the concentration of tamoxifen in a dose-dependent manner starting from slight change at 6.25µg/mL (57.2%) to maximum increase

at 12.5µg/mL (62.95%), as compared with the control. The increase in G1 phase was accompanied with no significant differences in S phase at 6.25µg/mL except at 12.5 and 25µg/mL that shown in table (1). On the other hand an apparent decreased G2/M phase population was observed in a concentration dependent manner after 24 hrs in comparison to the control. Induction of cell cycle arrest and apoptosis considered as the most important targets for developing anti-cancer drugs. Arresting the tumor cell cycle will dramatically inhibit the cell proliferation and resulting in programmed cell death. The resulted flow cytometric analysis revealed that tamoxifen cause accumulation of MCF-7 cells at G1 phase in a dose-dependent pattern which suggested that the transition from G1 to S phase was blocked by tamoxifen which was mainly observed at 12.5µg/mL this results agrees with Robert L. Sutherland when used Flow cytometric cell cycle analysis of tamoxifen-treated cultures showed an increase in the proportion of cells in the G1 phase of the cell cycle and also cells were not only arrested in the G1 phase but also in the G2 phase of the cell cycle (18). This study revealed that tamoxifen is a potent antagonist of estrogen and induces apoptosis in ER+ cells; however at concentrations, higher than 12.5µg/ml the level of G1 phase was decreased. At 25µg/mL concentration, of tamoxifen, showed less effect, on cell cycle, indicating, that the estrogen reversible, effect of tamoxifen, in MCF-7 cells, is mediated, via the estrogen receptor mechanism,, these, agreement, with Lykkesfeldt, AE indicating, that in high, concentration of, tamoxifen the estrogen, reversible, effect, of it in MCF-7 cells, is mediated, via the estrogen, receptor, mechanism (19).

Conclusion

Flow cytometric, cell, cycle analysis, of tamoxifen, cultures showed, an increase, in the proportion, of cells, in the G1 phase, of the cell cycle, (but not in very, high concentration). Tamoxifen, from, flow cytometric, results; showed, cell cycle, arrest toward, MCF-7. It, showed, dose-dependent, manner the maximum, concentration, at (12.5µg/mL), that more, effected, on G1 phase, accumulation, in MCF-7 cell cycle.

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